

7-Substituted-melatonin and 7-substituted-1-methylmelatonin analogues: Effect of substituents on potency and binding affinity

Rüdiger Faust,^a Peter J. Garratt,^a Maria Angeles Trujillo Pérez,^a Vincent J.-D. Piccio,^a
Christian Madsen,^b Ane Stenstrøm,^b Bente Frølund,^b Kathryn Davidson,^c
Muy-Teck Teh^c and David Sugden^{c,*}

^aDepartment of Chemistry, University College London, 20 Gordon Street, London WC1H 0AJ, UK

^bDepartment of Medicinal Chemistry, The Danish University of Pharmaceutical Sciences, DK 2100 Copenhagen, Denmark

^cDivision of Reproduction and Endocrinology, School of Biomedical and Health Sciences, Room 2.12N Hodgkin Building, King's College London, London SE1 1UL, UK

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Abstract—A series of 7-substituted melatonin and 1-methylmelatonin analogues were prepared and tested against human and amphibian melatonin receptors. 7-Substituents reduced the agonist potency of all the analogues in the *Xenopus laevis* melanophore assay, 7-bromomelatonin (**5d**) and *N*-butanoyl 7-bromo-5-methoxytryptamine (**5f**) being the most active compounds, but both were 42-fold less potent than melatonin (**1**). Whereas all the analogues bind with lower affinity at the human MT₁ receptor than melatonin, **5d**, **5f** and *N*-propanoyl 7-bromo-5-methoxytryptamine (**5e**) show a similar binding affinity to melatonin at the MT₂ receptor and consequently show some MT₂ selectivity. These results suggest that the receptor pocket around C-7 favours binding by an electronegative group, suggesting an electropositive region in this area of the receptor.

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1. Introduction

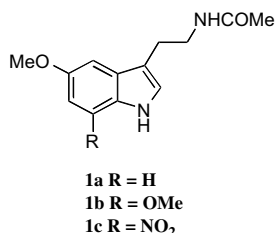
Melatonin (*N*-acetyl 5-methoxytryptamine, **1a**), the hormone secreted by the pineal gland, is a major component in the control of circadian rhythms in humans and other animals.^{1,2} Since the identification of three distinct melatonin receptor subtypes^{3–7} there have been numerous studies in an attempt to find analogues of melatonin that can distinguish between them.^{8–15} Such selective agents would allow both the assignment of the different functions of melatonin to specific receptors and provide models for the synthesis of compounds to target receptor subtypes. It has been found that the human MT₂ receptor subtype is much more tolerant to modification of the melatonin structure than the MT₁ receptor and a number of MT₂ selective compounds have been identified. Some progress has been made towards finding MT₁ selective compounds, but highly selective compounds have not yet been prepared.¹⁶ It has recently been suggested that melatonin receptor heterodimers

may be biologically significant which, while providing another potential target as an MT₁, MT₂ combination, may render the interpretation of *in vivo* findings more difficult.¹⁷

As part of our examination of the melatonin binding sites by preparing analogues of melatonin that have been altered at defined regions of the molecule, we have synthesised a variety of tryptamine derivatives substituted at position seven. It has been suggested from CoMFA model studies that there will be a low steric tolerance to substituents at positions 1, 6 and 7 of melatonin¹⁸ but few 7-substituted derivatives are known. 7-Methoxymelatonin (**1b**)¹⁹ and 7-nitromelatonin (**1c**)²⁰ have been prepared, the latter binding with 400-fold less affinity than melatonin at the MT₁ and 94-fold less at the MT₂ receptor. Because of our previous findings that the behaviour of β -substituted tryptamines was substantially modified by methyl substitution at *N*-1,²¹ we also prepared a similar series of 7-substituted-1-methyl compounds. The binding affinity of these compounds at the human MT₁ and MT₂ receptors was determined and their biological activity examined by the pigment aggregation response using a clonal *Xenopus laevis* melanophore cell line.

Keywords: Melatonin; Human melatonin receptors; Melanophores; Suzuki coupling reaction.

* Corresponding author. Tel.: +44 20 7848 6274; fax: +44 20 7848 6280; e-mail: david.sugden@kcl.ac.uk



2. Results and discussion

2.1. Chemistry

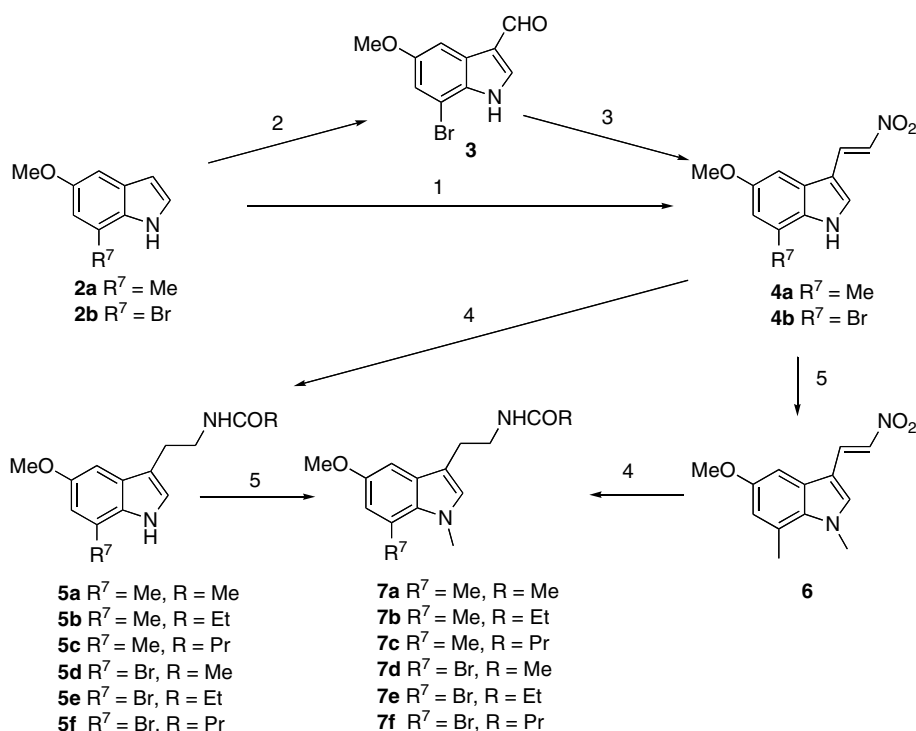
7-Methylmelatonin (**5a**), 7-bromomelatonin (**5d**),²² 1,7-dimethylmelatonin (**7a**), 7-bromo-1-methylmelatonin (**7d**) and their analogues were prepared as shown in Scheme 1. 5-Methoxy-7-methylindole (**2a**) was treated with 1-dimethylamino-2-nitroethylene by the method of Büchi and Mak²³ to give **4a** in 65% yield. The nitrovinyl derivative **4a** was then reduced to the saturated amine with LiAlH₄ and this was then directly converted to the amides **5a–c** by reaction with the appropriate acid chloride. This route proved less satisfactory with the 7-bromo derivatives and the two-step Vilsmeier–Henry procedure was found to be preferable. 7-Bromo-5-methoxyindole (**2b**)²⁴ was converted to the aldehyde **3** with POCl₃ and DMF and reaction of **3** with nitromethane and NH₄OAc gave **4b** in 50% overall yield. The nitrovinyl

derivative **4b** was then treated as for **4a** to give after reaction with the appropriate acid anhydride the amides **5d–f**. The 1,7-dimethyl derivatives **7a–c** were synthesised by methylation of **4a** with iodomethane and NaH to give **6**, which was then reduced and acylated by the same procedure used with **4a**. Attempts to methylate **4b** by the same procedure gave a mixture of the desired 7-bromo-5-methoxy-1-methyl-3-(2-nitrovinyl)indole and the corresponding debrominated derivative, so the amides **7d–f** were prepared by methylation of the amides **5d–f**.

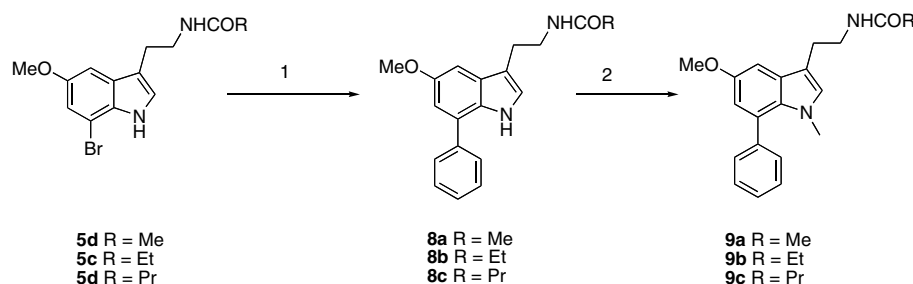
7-Bromo-5-methoxyindole (**2b**) and the acyl 7-bromotryptamines **5** and **7** are valuable intermediates for the synthesis of both 7-substituted indoles and related polycyclic systems. Treatment of 7-bromomelatonin (**5d**) with phenylboronic acid in a Suzuki coupling reaction²⁵ gave 7-phenylmelatonin (**8a**) in 17% yield, and treatment of **8a** with sodium hydride and MeI gave 1-methyl-7-phenylmelatonin (**9a**) in 14% yield. Similarly, **5e** and **5f** gave **8b** and **8c**, which could be converted to **9b** and **9c**, respectively (Scheme 2). 7-Phenylmelatonin (**8a**) was also prepared in similar overall yield from 7-bromo-5-methoxyindole by Suzuki coupling with phenylboronic acid followed by elaboration of the *N*-acetyl 3-ethylamino side chain.

2.2. Pharmacology

Table 1 shows the pK_i values for the binding affinity of melatonin and the 7-substituted and 7-substituted-1-methyl analogues on human MT₁ and MT₂ receptors and their potency (EC₅₀) on *Xenopus* melanophores.



Scheme 1. Reagents: (1) Me₂NCHCHNO₂, TFA; (2) a—POCl₃, DMF, CH₂Cl₂; b—NaOAc; (3) CH₃NO₂, THF, NH₄OAc; (4) LiAlH₄, THF, then RCOCl, Et₃N; (5) NaH, MeI, DMF.



Scheme 2. Reagents: (1) $C_6H_5B(OH)_2$, $Pd(PPh_3)_2Cl_2$, K_2CO_3 ; (2) NaH, MeI, DMF.

Table 1.

Compound	R ₁	R ₇	R	Melanophores pEC ₅₀	MT ₁ pK _i	MT ₂ pK _i	MT ₂ /MT ₁
1a melatonin	H	H	Me	10.10 ± 0.03	9.58 ± 0.04	9.47 ± 0.01	0.8
5a	H	Me	Me	7.34 ± 0.02	8.57 ± 0.05	8.76 ± 0.01	1.5
5b	H	Me	Et	7.50 ± 0.02	8.75 ± 0.09	8.96 ± 0.09	1.6
5c	H	Me	Pr	7.90 ± 0.01	8.54 ± 0.06	9.17 ± 0.07	4.3
7a	Me	Me	Me	8.01 ± 0.01	8.06 ± 0.05	8.81 ± 0.08	5.6
7b	Me	Me	Et	7.80 ± 0.03	7.80 ± 0.09	8.58 ± 0.08	6
7c	Me	Me	Pr	7.78 ± 0.04	7.71 ± 0.03	8.87 ± 0.04	14.5
5d	H	Br	Me	8.48 ± 0.02	8.45 ± 0.10	9.17 ± 0.05	5.2
5e	H	Br	Et	7.94 ± 0.01	8.38 ± 0.04	9.27 ± 0.02	7.8
5f	H	Br	Pr	8.46 ± 0.02	8.29 ± 0.02	9.45 ± 0.03	14.5
7d	Me	Br	Me	7.61 ± 0.05	7.98 ± 0.13	8.88 ± 0.02	7.9
7e	Me	Br	Et	7.19 ± 0.01	7.85 ± 0.04	8.64 ± 0.02	6.2
7f	Me	Br	Pr	7.28 ± 0.04	7.48 ± 0.09	8.67 ± 0.04	15.5
8a	H	Ph	Me	6.92 ± 0.01	4.89 ± 0.54	5.52 ± 0.26	4
8b	H	Ph	Et	6.81 ± 0.07	6.21 ± 0.02	6.21 ± 0.02	1
8c	H	Ph	Pr	7.34 ± 0.03	6.02 ± 0.08	6.06 ± 0.12	1
9a	Me	Ph	Me	6.47 ± 0.02	5.90 ± 0.02	6.34 ± 0.10	2.8
9b	Me	Ph	Et	NA	5.23 ± 0.01	6.17 ± 0.02	8.7
9c	Me	Ph	Pr	NA	5.27 ± 0.02	6.07 ± 0.04	6.3

MT₁ and MT₂ receptor binding data are means ± SEM of quadruplicate determinations. Agonist data on melanophores are means ± SEM of triplicate experiments. NA, no agonist effect detected at 10 μM.

NA, no agonist effect detected at 10 μM.

2.3. Discussion

The effects of introducing a methyl, bromine or phenyl group at 7 are shown in the Table 1. In the case of melanophores, the introduction of any of these substituents drastically reduces the potency, with **5d** and **5f** the most potent compounds and **9a** the least, **9b** and **9c** showing no activity. 7-Methylmelatonin (**5a**) is 575 times, 7-bromomelatonin (**5d**) 42 times and 7-phenylmelatonin (**8a**) 1500 times less potent than melatonin. The greater potency of the 7-bromo compounds over their 7-methyl analogues suggests that there may be an electrostatic binding effect from the bromine atom. The 7-phenyl derivatives are the least active, probably because of the greater steric interaction of the phenyl group than that of methyl or bromine. The considerable decrease in potency of the 7-substituted derivatives is in striking contrast to their binding affinity to the

human MT₂ receptor. All three 7-bromo analogues have high binding affinities at MT₂, increasing with increasing size of the *N*-acyl group, whereas they show a somewhat lower affinity for the MT₁ receptor and the affinity decreases with increasing alkyl group size. Thus, *N*-butanoyl 7-bromo-5-methoxytryptamine (**5f**) has the same binding affinity at the MT₂ receptor as melatonin, although it binds much more weakly at the MT₁ receptor (MT₂/MT₁ ratio 14:5). 7-Methylmelatonin (**5a**) binds only 5 times less strongly to MT₂ than melatonin, although it is 575 times less active on melanophores, and the difference in binding between MT₁ and MT₂ is not large (MT₂/MT₁ ratio 1:5). The previously reported 7-nitromelatonin (**1c**)¹⁸ has reported binding affinities more closely resembling those of the 7-bromo derivatives, with an MT₂/MT₁ selectivity of 4, but binds significantly less strongly to MT₂ than does **5a** (Fig. 1).

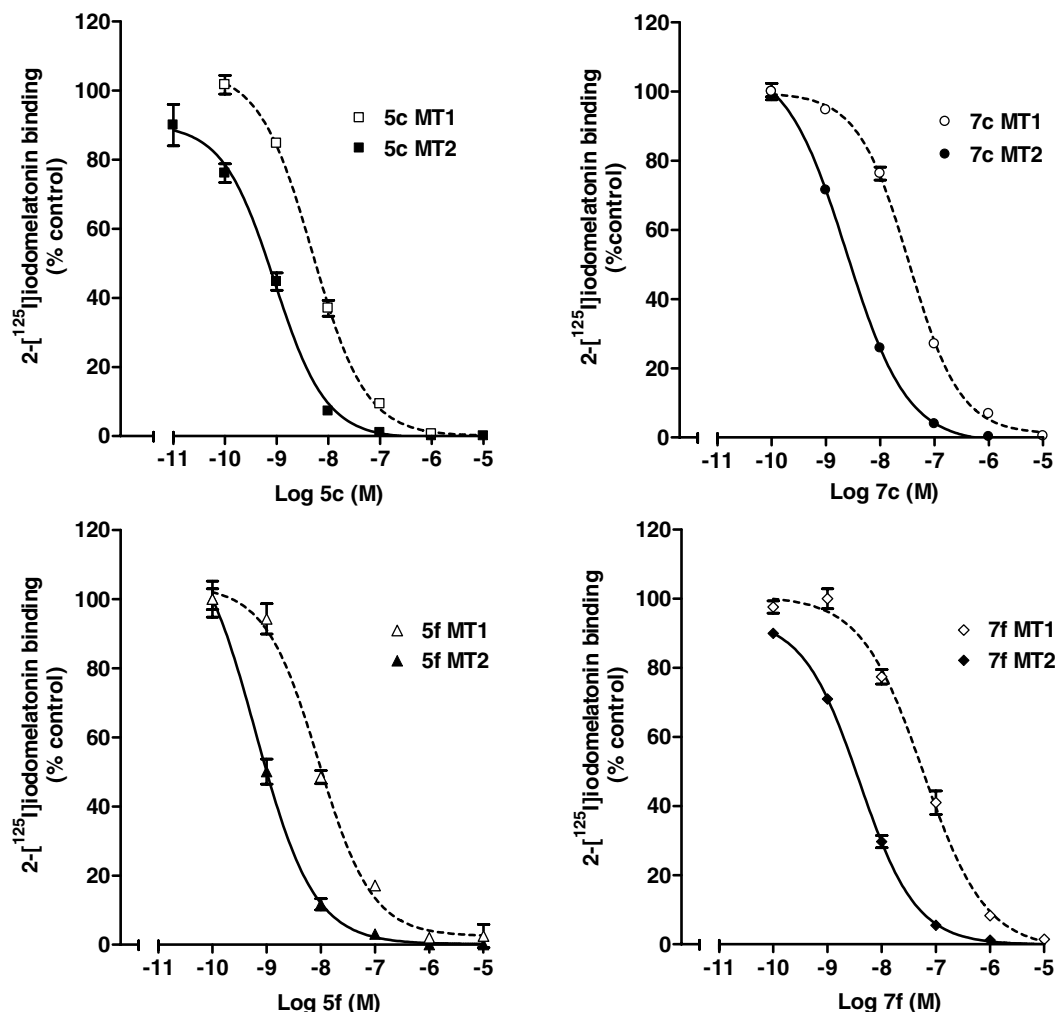


Figure 1. Competition radioligand binding curves for compounds **5c**, **7c**, **5f** and **7f** on recombinant human MT₁ and MT₂ receptor subtypes. Each data point represents the mean \pm SEM of triplicate samples.

Introduction of the N-1 methyl group in the 7-methyl series has a rather small effect, decreasing the binding affinity to MT₁, but having little effect on potency at melanophores. In the 7-bromo and 7-phenyl series there is generally a reduction in both binding and potency, with *N*-propanoyl 5-methoxy-1-methyl-7-phenyltryptamine (**9b**) and the *N*-butanoyl analogue **9c** being inactive on melanophores but binding weakly to the human receptors. All of the 7-substituted derivatives show a higher binding affinity for MT₂ over MT₁ except for the 7-phenyl derivatives **8b** and **8c**, which are equipotent. The behaviour of compound **8a** is somewhat anomalous in that it shows lower binding affinity to MT₁ and MT₂ than would have been expected from its affinity to melanophores and it also has less binding affinity than the corresponding *N*-methyl derivative **9a**.

The 7-methyl analogue **7c** (N¹Me, R = Pr) and the 7-bromo analogues **5f** (N¹H, R = Pr) and **7f** (N¹Me, R = Pr) may provide leads to compounds with differing activities at the human MT₁ and MT₂ receptors. Although these compounds have a lower binding affinity than melatonin, they already show some preference for binding to the MT₂ receptor and β -substitution, for

example, may increase both the binding affinity and the discrimination between receptors.²¹ Since 7-bromomelatonin shows a higher binding affinity than 7-methylmelatonin, β -substituted 7-fluoromelatonin analogues may be the targets of choice. We are currently exploring these concepts.

3. Experimental

3.1. Chemistry

Melting points were determined either in glass capillary tubes on a Gallenkamp Melting Point Apparatus or a Reichert-Jung Thermovar hot-stage and are uncorrected. EI mass spectra were recorded on a VG ZAB-2F, CI mass spectra on a VG 12-250, FAB on a M550 and GCMS on a Shimadzu QP5050 mass spectrometer. Only molecular (M⁺) or (M+1)⁺ ions and the most significant peaks are reported. ¹H NMR spectra were taken on Varian VXR-400, Varian Gemini 2000, Varian Mercury, or Bruker AC300 spectrometer in CDCl₃ unless stated otherwise and are reported in δ . ¹³C NMR spectra were taken in CDCl₃, unless stated otherwise, and are

reported in δ . Solvents were dried over sodium where appropriate or over activated sieves. Chromatography was carried out on Sorbsil c60-silica or Merck silica gel. Analytical TLC was carried out on DC-Alufolien plates visualised with ultraviolet light, iodine, potassium permanganate or ninhydrin. Microanalyses were performed by the Microanalytical Section, Department of Chemistry, UCL, or the Analytic Research Department, H. Lundbeck A/S, Denmark. Satisfactory analyses or high resolution mass spectra data were obtained for all new analogues.

3.1.1. 5-Methoxy-7-methyl-3-(2-nitrovinyl)indole (4a).

Trifluoroacetic acid (TFA) (1.82 g, 16 mmol) in CH_2Cl_2 (4 mL) was added to a suspension of 1-dimethylamino-2-nitroethylene (0.95 g, 8.2 mmol) in CH_2Cl_2 (5 mL) at 0 °C. A solution of 5-methoxy-7-methylindole²⁶ (2a, 0.89 g, 5.5 mol) in CH_2Cl_2 (5 mL) was then added and the mixture stirred at 0 °C for 30 min and then allowed to come to room temperature and stirred for 5 h. Ice (5 g) was added and the mixture was separated and the aqueous layer extracted with CH_2Cl_2 (3× 25 mL). The combined organic layers were washed with a saturated solution of NaHCO_3 (3× 20 mL) and dried (MgSO_4). The solvent was removed under reduced pressure to give 4a as an orange solid, 0.83 g (3.6 mmol, 65%), mp 225–226 °C; ^1H NMR 3.89 (s, 3H), 6.77 (d, $J = 2.0$ Hz, 1H), 7.25 (d, $J = 2.2$ Hz, 1H), 7.86 (d, $J = 13.4$ Hz, 1H), 8.08 (d, $J = 2.9$ Hz, 1H), 8.35 (d, $J = 13.4$ Hz, 1H), 11.20 (br s, 1H); ^{13}C NMR δ 16.9, 110.2, 118.2, 121.5, 122.8, 124.9, 131.9, 131.8, 133.0, 133.5, 136.0, 138.0; MS *m/e* 202 (100), 155, 129. Found: 203.0820. $\text{C}_{11}\text{H}_{11}\text{N}_2\text{O}_2$ ($\text{M}^+ + 1$) requires 203.0815.

3.2. General method for the preparation of amides (5a–c)

A solution of 4a (0.20 g, 0.86 mmol) in THF (5 mL) was added dropwise to a stirred suspension of LiAlH_4 (0.19 g, 5.16 mmol) in THF (10 mL) at 0 °C. The mixture was allowed to warm to room temperature and stirring was continued for 18 h. Water (3 mL), aqueous NaOH (2 M, 2.2 mL) and ether (6 mL) were then added sequentially and the mixture was filtered and the precipitate washed with water and THF. The combined filtrate was separated and the aqueous layer extracted with ether (3× 25 mL). The combined organic layers were washed with brine (25 mL) and dried (MgSO_4). A stirred solution of the crude tryptamine (0.16 g, 0.80 mmol) in CH_2Cl_2 at 0 °C was then treated with the appropriate acid chloride (0.80 mmol), the solution allowed to warm to room temperature and stirring continued for 4 h. The solution was washed with aqueous HCl (10%, 25 mL), saturated NHCO_3 (25 mL) and brine (25 mL) and the organic phase dried (MgSO_4). The solvent was evaporated under reduced pressure and the brown oil purified by flash chromatography on silica gel, eluting with ether/acetone, to give the amide.

3.2.1. 7-Methylmelatonin (N-acetyl 5-methoxy-7-methyl-tryptamine) (5a). White solid, 0.40 g (29% from 4a) mp 125–126 °C; ^1H NMR δ 1.90 (s, 3H), 2.43 (s, 3H), 2.91 (t, $J = 6.5$ Hz, 2H), 3.56 (d, $J = 6.5$ Hz, 2H), 3.82 (s, 3H), 5.57 (s, 1H), 6.68 (d, $J = 1.2$ Hz, 1H), 6.86 (d,

$J = 2.0$ Hz, 1H), 6.99 (d, $J = 2.0$ Hz, 1H), 8.03 (s, 1H); ^{13}C NMR δ 16.6, 23.3, 25.3, 39.7, 55.8, 97.8, 105.0, 113.0, 121.5, 122.3, 127.0, 131.1, 154.2, 170.0, 177.8; MS *m/e* 246 (100), 187, 174. Anal. ($\text{C}_{12}\text{H}_{12}\text{N}_2\text{O}$) C, H, N.

3.2.2. N-Propanoyl 5-methoxy-7-methyltryptamine (5b).

Brown oil, 0.60 g (41% from 4a); ^1H NMR δ 1.09 (t, $J = 7.6$ Hz, 3H), 2.43 (s, 3H), 2.91 (t, $J = 6.6$ Hz, 2H), 3.46 (d, $J = 7.0$ Hz, 2H), 3.57 (d, $J = 6.5$ Hz, 2H), 3.82 (s, 3H), 5.48 (s, 1H), 6.68 (d, $J = 1.4$ Hz, 1H), 6.86 (d, $J = 2.3$ Hz, 1H), 7.00 (d, $J = 2.3$ Hz, 1H), 7.86 (s, 1H); MS *m/e* 260 (100), 187, 174. Found: 261.1603. $\text{C}_{15}\text{H}_{21}\text{O}_2\text{N}_2$ ($\text{M}^+ + 1$) requires 261.1610.

3.2.3. N-Butanoyl 5-methoxy-7-methyltryptamine (5c).

Brown oil, 0.33 g 22% from 4a: ^1H NMR δ 0.89 (t, $J = 7.3$ Hz, 3H), 1.60 (q, 7.3 Hz, 3H), 1.60 (m, 2H), 2.06 (t, 7.3 Hz, 2H), 2.43 (s, 3H), 2.91 (t, $J = 6.6$ Hz, 2H), 3.82 (s, 3H), 5.48 (s, 1H), 6.68 (d, $J = 1.4$ Hz, 1H), 6.86 (d, $J = 2.0$ Hz, 1H), 7.00 (d, $J = 2.0$ Hz, 1H), 7.85 (s, 1H); ^{13}C NMR δ 13.7, 16.6, 19.1, 25.4, 38.8, 39.4, 55.8, 97.9, 113.3, 121.5, 122.3, 127.1, 131.2, 154.3, 164.5, 172.9; MS *m/e* 274 (100), 187, 174. Found: 275.1760. $\text{C}_{16}\text{H}_{23}\text{O}_2\text{N}_2$ ($\text{M}^+ + 1$) requires 275.1750.

3.2.4. 7-Bromo-3-formyl-5-methoxyindole (3).

7-Bromo-5-methoxyindole²⁷ in CH_2Cl_2 was added dropwise to a cold (0–5 °C), stirred mixture of POCl_3 (0.30 g, 1.95 mmol) and DMF (0.14 g, 1.95 mmol) in CH_2Cl_2 under N_2 . The reaction mixture was allowed to warm to room temperature and then refluxed for 2 h. After cooling, aqueous NaOAc (8 mL, 2 M) was added and the vigorously stirred mixture refluxed for 15 min. After cooling, CH_2Cl_2 (10 mL) and water (10 mL) were added, the mixture was separated and the aqueous phase extracted with CH_2Cl_2 (3× 10 mL). The combined organic extracts were washed with saturated Na_2CO_3 solution (3× 15 mL), brine (20 mL) and dried (MgSO_4). The solvent was removed under vacuo and the residue chromatographed on silica gel, eluting with EtOAc/petroleum ether 800–100 (2:1), to give 3 as pale yellow crystals, 0.30 g, 67%, mp 166–168 °C; ^1H NMR ($\text{DMSO}-d_6$) δ 3.80 (s, 3H), 7.14 (d, $J = 2.2$ Hz, 1H), 7.58 (d, $J = 2.2$ Hz, 1H), 8.29 (d, $J = 2.75$ Hz, 1H), 9.90 (s, 1H), 12.26 (br s, 1H); ^{13}C NMR 55.6, 102.2, 104.8, 115.3, 118.5, 125.8, 130.3, 139.1, 155.8, 184.9. Anal. ($\text{C}_{10}\text{H}_8\text{BrNO}_2$) C, H, N.

3.2.5. 7-Bromo-5-methoxy-3-(2-nitrovinyl)indole (4b).

The aldehyde 3 (0.20 g, 0.79 mmol) and NH_4Ac (0.066 g, 0.87 mmol), MeNO_2 (47 μL , 0.87 mmol) were dissolved in THF (3 mL) and refluxed for 20 h. The mixture was cooled to room temperature and water (6 mL) was added. The aqueous layer was extracted with EtOAc (3× 10 mL), the organic phases combined and washed with saturated NaHCO_3 (2 mL), brine (5 mL) and dried (MgSO_4). The solvent was removed under vacuo and the residue purified by column chromatography (SiO_2 , petrol/EtOAc, 50:50) to give 4b (174 mg, 0.59 mmol, 75%), as yellow crystals, mp 209–211 °C (dec); ^1H NMR ($\text{DMSO}-d_6$) δ 3.86 (s, 3H), 7.13 (d, $J = 2.2$ Hz, 1H), 7.45 (d, $J = 2.2$ Hz, 1H), 8.09 (d,

$J = 13.2$ Hz, 1H), 8.26 (d, $J = 2.75$ Hz, 1H), 8.31 (d, $J = 13.2$ Hz, 1H), 12.28 (br s, 1H).

3.2.6. 7-Bromomelatonin (*N*-acetyl 7-bromo-5-methoxytryptamine) (5d). A solution of **4b** (0.15 g, 0.50 mmol) in THF (2.5 mL) was added to a stirred suspension of LiAlH_4 (0.09 g, 3.6 mmol) in THF (4 mL) under N_2 at 0°C . The mixture was allowed to warm to room temperature and stirred for 1 h. The mixture was poured into ice-cold NaOH solution (2 mL, 1 M) and water (1.0 mL) and ether (15 mL) were added and the mixture filtered. Ether (15 mL) was added and the solution dried over MgSO_4 . The solvent was removed under vacuo and the residue (0.19 g) was dissolved in dry THF (3 mL) at 0°C and Et_3N (0.15 mL, 1.1 mmol) was added and the mixture stirred for 5 min before the addition of acetic anhydride (0.10 mL, 1.1 mmol). The mixture was allowed to warm to room temperature and stirred for 2 h. It was then extracted with HCl (2 \times 25 mL, 2 M), saturated NaHCO_3 (50 mL), brine (50 mL) and dried (MgSO_4). Chromatography on silica gel, eluting with EtOAc/petroleum ether (3:1), gave **5d**, 0.072 g (0.21 mmol, 42%) as white crystals, mp 141–143 $^\circ\text{C}$; ^1H NMR (CD_3OD) δ 1.91 (s, 3H), 2.87 (t, $J = 7.2$ Hz, 2H), 3.40–3.47 (m, 2H), 3.81 (s, 3H), 6.94 (d, $J = 2.2$ Hz, 1H), 7.05 (d, $J = 2.2$ Hz), 7.10 (s, 1H), 8.04 (br s, 1H), 10.32 (br s, 1H); ^{13}C NMR (CD_3OD) δ 22.7, 26.3, 41.4, 56.5, 100.9, 105.4, 114.3, 114.9, 125.3, 130.0, 131.8, 155.2, 173.1. Anal. ($\text{C}_{13}\text{H}_{15}\text{BrN}_2\text{O}_2$) C, H, N.

3.2.7. *N*-Propanoyl 7-bromo-5-methoxytryptamine (5e). Prepared from **4b** as for **5d**, acylating with propionic anhydride; 0.042 g (0.130 mmol, 62%), off-white crystals, mp 95 $^\circ\text{C}$ (EtOAc); ^1H NMR δ 1.00 (t, $J = 7.5$ Hz, 3H), 2.04 (q, $J = 7.6$ Hz, 2H), 2.76 (t, $J = 7.2$ Hz, 2H), 3.29 (q, $J = 7.0$ Hz, 2H), 3.77 (s, 3H), 6.96 (d, $J = 2.1$ Hz, 1H), 7.06 (d, $J = 2.1$ Hz, 1H), 7.16 (d, $J = 2.2$ Hz, 1H), 7.83 (bt, 1H), 10.86 (s, 1H); ^{13}C NMR (DMSO) 13.6, 18.6, 25.1, 37.4, 39.0, 39.7, 55.7, 75.5, 100.3, 113.1, 124.8; MS *m/e* 326, 324, 253, 251, 240, 238. Anal. ($\text{C}_{14}\text{H}_{17}\text{BrN}_2\text{O}_2$) C, H, N.

3.2.8. *N*-Butanoyl 7-bromo-5-methoxytryptamine (5f). Prepared from **4b** as for **5d**, acylating with butyric anhydride; 0.044 g (0.130 mmol, 62%), off-white crystals, mp 87 $^\circ\text{C}$ (EtOAc); ^1H NMR δ 0.81 (t, $J = 7.4$ Hz, 3H), 1.48 (m, 2H), 2.01 (t, $J = 7.3$ Hz, 2H), 2.75 (t, $J = 7.2$ Hz, 2H), 3.30 (q, $J = 7.2$ Hz, 2H), 3.76 (s, 3H), 6.95 (d, $J = 2.2$ Hz, 1H), 7.06 (d, $J = 2.1$, 1H), 7.15 (d, $J = 2.3$ Hz, 1H), 7.85 (bt, 1H), 10.86 (s, 1H); ^{13}C NMR (DMSO) δ 13.6, 18.6, 25.15, 37.4, 55.75, 100.3, 104.0, 113.1, 124.8, 128.7, 129.1, 153.3, 171.8; MS *m/e* 342, 340, 253, 251, 240, 238. Anal. ($\text{C}_{15}\text{H}_{19}\text{BrN}_2\text{O}_2$) C, H, N.

3.2.9. 1,7-Dimethyl-5-methoxy-3-(2-nitrovinyl)indole (6). Sodium hydride (1 mmol, 55–65% in oil) was added to a stirred solution of **4a** (0.20 g, 0.86 mmol) in DMF (5 mL) at 0°C . After 30 min iodomethane (0.14 g, 1.0 mmol) was added and the mixture allowed to warm to room temperature and stirred for 16 h. The mixture was poured into water (10 mL), the solution brought

to pH 7 by addition of aqueous HCl (2 M) and allowed to stand for 4 h at 4°C . The orange precipitate was removed by filtration, dried under vacuo and purified by chromatography on silica gel, eluting with CH_2Cl_2 /acetone, to give **6**, 0.11 g (0.45 mmol, 52%); ^1H NMR δ 2.69 (s, 3H), 3.85 (s, 3H), 4.05 (s, 3H), 6.68 (d, $J = 1.9$ Hz, 1H), 6.86 (d, $J = 1.9$ Hz, 1H), 7.34 (s, 1H), 8.63 (d, $J = 13.4$ Hz, 1H), 8.20 (d, $J = 13.4$ Hz, 1H); ^{13}C NMR δ 19.5, 37.8, 55.7, 100.1, 107.4, 116.4, 123.8, 127.7, 131.3, 132.0, 133.4, 137.8, 155.9; MS *m/e* 246 (100), 203, 188.

3.2.10. General method for the preparation of amides (7a–c). A solution of **6** (0.20 g, 0.86 mmol) in THF (5 mL) was added dropwise to a stirred suspension of LiAlH_4 (0.19 g, 5.16 mmol) in THF (10 mL) at 0°C . The solution was allowed to warm to room temperature and stirred for 18 h. Water (3 mL), aqueous NaOH (2.2 mL, 2 M) and ether (6 mL) were added sequentially and the mixture was filtered. The residue was washed with THF (5 mL) and ether (5 mL) and the combined filtrates were separated and the aqueous layer extracted with ether (3 \times 25 mL). The combined organic layers were washed with brine (25 mL) and dried (MgSO_4). Removal of the solvent under reduced pressure gave the amine, which was used without further purification. The crude amine (0.80 mmol) was dissolved in CH_2Cl_2 (5 mL), the stirred solution cooled to 0°C and Et_3N (0.10 mL, 0.77 mmol) and the appropriate acid chloride (0.80 mmol) added. The mixture was allowed to warm to room temperature and stirred for 4 h. The solution was then extracted with aqueous HCl (5 mL, 10%), satd NaHCO_3 solution (5 mL), and brine (5 mL). The organic phase was dried (MgSO_4), the solvent removed under reduced pressure and the residue purified by flash chromatography on silica gel, eluting with ether/acetone, to give the desired amide.

3.2.11. 1,7-Dimethylmelatonin (*N*-acetyl 5-methoxy-1,7-dimethyltryptamine) (7a). White solid, 0.10 g (0.38 mmol, 44%), mp 131–133 $^\circ\text{C}$; ^1H NMR δ 2.42 (s, 3H), 3.19 (s, 3H), 3.37 (t, $J = 6.6$ Hz, 2H), 4.04 (d, $J = 6.6$ Hz, 2H), 4.32 (s, 3H), 4.46 (s, 3H), 6.07 (s, 1H), 7.09 (d, $J = 2.2$ Hz, 1H), 7.31 (d, $J = 2.2$ Hz, 1H), 7.75 (s, 1H); ^{13}C NMR δ 19.5, 23.4, 25.0, 36.5, 39.7, 55.7, 97.8, 110.6, 114.7, 122.6, 128.9, 129.1, 131.1, 153.6, 169.9; MS *m/e* 260, 201, 188, 173. $\text{C}_{15}\text{H}_{20}\text{O}_2\text{N}_2$ requires 260.1510. Found: 260.1525.

3.2.12. *N*-Propanoyl 5-methoxy-1,7-dimethyltryptamine (7b). White solid, 0.11 g (0.40 mmol, 45%), mp 107–109 $^\circ\text{C}$ (EtOAc); ^1H NMR δ 1.62 (t, $J = 7.5$ Hz, 2H), 2.64 (q, $J = 7.5$ Hz, 2H), 3.20 (s, 3H), 3.38 (t, $J = 6.6$ Hz, 2H), 4.05 (q, $J = 6.6$ Hz, 2H), 4.33 (s, 3H), 4.47 (s, 3H), 6.04 (s, 1H), 7.10 (d, $J = 2.5$ Hz, 1H), 7.32 (d, $J = 2.5$ Hz, 1H) 7.76 (s, 1H); ^{13}C NMR δ 9.8, 19.5, 25.1, 29.8, 36.5, 39.6, 55.7, 97.8, 110.7, 114.7, 122.6, 128.9, 129.1, 131.1, 153.6, 173.6; MS *m/e* 274, 201, 188, 173. Anal. ($\text{C}_{16}\text{H}_{22}\text{N}_2\text{O}_2$) C, H, N.

3.2.13. *N*-Butanoyl 5-methoxy-1,7-dimethyltryptamine (7c). White solid, 0.16 g (0.56 mmol, 65%), mp 106–107 $^\circ\text{C}$; ^1H NMR δ 0.89 (t, $J = 7.4$ Hz, 3H), 1.61 (m,

2H), 2.07 (t, $J = 7.4$ Hz, 2H), 2.68 (s, 3H), 2.85 (t, $J = 6.6$ Hz, 2H), 3.54 (q, $J = 6.6$ Hz, 2H), 3.80 (s, 3H), 3.95 (s, 3H), 5.49 (s, 1H), 6.58 (d, $J = 2.0$ Hz, 1H), 6.71 (s, 1H), 6.79 (d, $J = 2.0$ Hz, 1H); ^{13}C NMR δ 13.7, 19.1, 19.5, 25.1, 36.5, 38.8, 39.5, 55.7, 97.8, 110.7, 114.7, 122.6, 128.9, 129.1, 131.1, 153.6, 172.8; MS *m/e* 288, 201, 188, 173. Found: 288.1838. $\text{C}_{17}\text{H}_{24}\text{N}_2\text{O}_2$ requires 288.1850.

3.2.14. 7-Bromo-1-methylmelatonin (*N*-acetyl 7-bromo-5-methoxy-1-methyltryptamine) (7d). Sodium hydride (20 mg, 0.48 mmol) was added to a stirred solution of **5d** (0.070 g, 0.218 mmol) in dry THF (1.0 mL) at 0 °C and stirring continued for 30 min. Iodomethane (27 μL , 0.436 mmol) was then added, the mixture allowed to warm to room temperature and stirred for 5 h. Water (2.0 mL) and EtOAc (2 mL) were then added, the organic layer separated. The aqueous layer was extracted with EtOAc (3 \times 2 mL) and the combined organic fractions were washed with brine (2 mL), dried (MgSO_4) and concentrated under reduced pressure. The residue was purified by column chromatography (SiO_2 , EtOAc) and recrystallised from CH_2Cl_2 –petrol to give **7d** (0.010 g, 0.031 mmol, 14%); mp 109 °C; ^1H NMR (CD_2Cl_2) δ 1.88 (s, 3H), 2.85 (t, $J = 6.7$ Hz, 2H), 3.47 (m, $J = 6.7$ Hz, 2H), 3.81 (s, 3H), 4.05 (s, 3H), 5.55 (br s, 1H), 6.84 (s, 1H), 6.97 (d, $J = 2.2$ Hz, 1H), 7.01 (d, $J = 2.2$ Hz, 1H); ^{13}C NMR (CD_2Cl_2) δ 23.4, 25.3, 36.7, 39.9, 56.2, 100.5, 104.2, 111.4, 116.6, 129.4, 130.9, 131.2, 154.1, 169.9; MS *m/e* 326 (33, $\text{M}+2$), 324 (32, M^+), 252 (100). Found: M^+ 324.0481. $\text{C}_{14}\text{H}_{17}^{79}\text{BrN}_2\text{O}_2$ requires 324.0473.

3.2.15. *N*-Propanoyl 7-bromo-5-methoxy-1-methyltryptamine (7e). From **5e** by the same procedure as for **7d**. 0.028 g, 0.067 mmol, 67%; white crystals, mp 93 °C (CH_2Cl_2 , petrol); ^1H NMR (CD_2Cl_2) δ 1.08 (t, $J = 7.6$ Hz, 3H), 2.12 (q, $J = 7.6$ Hz, 2H), 2.85 (t, $J = 6.9$ Hz, 2H), 2.85 (t, $J = 6.9$ Hz, 2H), 3.48 (q, $J = 6.6$ Hz, 2H), 3.81 (s, 3H), 4.04 (s, 3H), 5.61 (br s, 1H), 6.83 (s, 1H), 6.97 (d, $J = 2.3$ Hz), 7.05 (d, $J = 2.3$ Hz, 1H); ^{13}C NMR (CD_2Cl_2) δ 10.0, 25.4, 30.0, 36.7, 39.9, 56.3, 100.6, 104.3, 111.5, 116.7, 129.4, 131.0, 131.3, 154.1, 173.7; MS *m/e* 340, 338, 265, 267, 252, 254. Found: 338.0632. $\text{C}_{15}\text{H}_{19}\text{N}_2\text{O}_2^{79}\text{Br}$ requires 338.0630.

3.2.16. *N*-Butanoyl 7-bromo-5-methoxy-1-methyltryptamine (7f). From **5f** by the same procedure as for **7d**. 0.012 g, 0.034 mmol, 55%; white crystals, mp 111 °C (EtOAc/petrol); ^1H NMR (DMSO) δ 0.82 (t, $J = 7.4$ Hz, 3H), 1.49 (m, 2H), 2.00 (t, 7.3 Hz, 2H), 2.72 (t, $J = 7.3$ Hz, 2H), 3.26 (q, $J = 6.8$ Hz, 2H), 3.76 (s, 3H), 3.97 (s, 3H), 6.94 (d, $J = 2.2$ Hz, 1H), 7.05 (d, $J = 2.2$ Hz, 1H), 7.11 (s, 1H), 7.86 (m, 1H); ^{13}C NMR (DMSO) δ 13.6, 18.6, 24.7, 35.9, 37.4, 38.6, 55.6, 100.5, 103.1, 111.1, 115.2, 128.1, 130.8, 131.1, 153.1, 171.8; MS *m/e* 355, 353, 275. Found: 353.0857. $\text{C}_{16}\text{H}_{22}\text{N}_2\text{O}_2^{79}\text{Br}$ (M^++1) requires 353.0865.

3.2.17. 7-Phenylmelatonin (*N*-acetyl 5-methoxy-7-phenyltryptamine) (8a). A solution of aqueous potassium carbonate (3 M, 0.3 mL) was added to a solution of 7-bromomelatonin **5d** (0.115 g, 0.37 mmol) and phenyl-

boronic acid (0.090 g, 0.74 mmol) in DMF (4.5 mL) was added. The resulting mixture was flushed with argon and $\text{Pd}(\text{PPh}_3)_2\text{Cl}_2$ (0.029 g, 0.04 mmol) was added rapidly. The mixture was stirred at 100 °C for 20 h and then allowed to cool to room temperature before being filtered through Celite. EtOAc (50 mL) was added to the filtrate and the organic phase separated, washed with water (50 mL), dried (MgSO_4) and concentrated under reduced pressure. The residue was purified by HPLC (Chiralpak AD, 20 \times 250, 10 Å, 10 mL min^{-1} , hexane/IPA 80:20, 254 nm, 1 mL loop, room temperature = 18.5 min) to give **8a** 0.017 g, (0.055 mmol, 15%), white solid, mp 62 °C; ^1H NMR (CD_2Cl_2) δ 1.89 (3H, s), 2.95 (t, $J = 6.7$ Hz, 2H), 3.55 (m, 2H), 3.88 (s, 3H), 5.61 (br s, 1H) 6.89 (d, $J = 2.4$ Hz, 1H), 7.06 (d, $J = 2.3$ Hz, 1H), 7.08 (d, $J = 2.4$ Hz, 1H) 7.41–7.65 (m, 5H), 8.25 (br s, 1H); ^{13}C δ (CD_2Cl_2) 24.9, 27.2, 41.5, 57.7, 101.6, 113.7, 115.0, 124.9, 128.2, 129.4, 129.9, 130.2, 130.9, 131.2, 140.5, 156.3, 171.5; MS *m/e* 308 (6%, M^+), 249, 235 86, 84 (100). Found: M^+ 308.1514. $\text{C}_{19}\text{H}_{20}\text{N}_2\text{O}_2$ requires 308.1525.

3.2.18. *N*-Propanoyl 5-methoxy-7-phenyltryptamine (8b). 0.052 g (0.161 mmol, 40%), white crystals, mp 48 °C; ^1H NMR (CD_2Cl_2) δ 1.08 ($J = 7.6$ Hz, 3H), 2.12 (q, $J = 7.6$ Hz, 2H), 2.94 (t, $J = 6.8$ Hz, 2H), 3.56 (q, $J = 6.5$ Hz, 2H), 3.88 (s, 3H), 5.61 (br s, 1H), 6.89 (d, $J = 2.3$ Hz, 1H), 7.06 (m, 2H), 7.43 (m, 1H), 7.52 (m, 2H), 7.65 (m, 2H), 8.30 (br s, 1H); ^{13}C NMR (CD_2Cl_2) 11.5, 27.25, 31.4, 41.4, 57.67, 101.7, 113.7, 115.0, 125.0, 128.2, 129.4, 129.9, 130.2, 130.9, 131.2, 140.6, 156.2, 175.3; MS *m/e* 322, 249 (100), 236. Found: 322.1683. $\text{C}_{20}\text{H}_{22}\text{N}_2\text{O}_2$ requires 322.1681.

3.2.19. *N*-Butanoyl 5-methoxy-7-phenyltryptamine (8c). 0.051g (0.151 mmol, 38%), colourless oil; ^1H NMR (CD_2Cl_2) δ 0.84 (t, $J = 5.5$ Hz, 3H), 1.53 (t, $J = 5.6$ Hz, 2H), 2.01 (t, $J = 5.6$ Hz, 2H), 2.88 (t, $J = 5.2$ Hz, 2H), 3.49 (q, $J = 4.9$ Hz, 2H), 3.82 (s, 3H), 5.67 (br s, 1H), 6.83 (d, $J = 1.6$ Hz, 1H), 6.99 (d, $J = 1.6$ Hz, 1H), 7.00 (d, $J = 1.5$ Hz, 1H), 7.36 (m, 1H), 7.42 (m, 1H), 7.57 (m, 2H), 8.39 (br s, 1H); ^{13}C NMR (CD_2Cl_2) δ 15.4, 21.0, 40.4, 41.3, 57.6, 101.7, 113.7, 125.0, 128.2, 129.4, 129.9, 130.2, 130.9, 131.2, 140.6, 156.2, 174.5; MS *m/e* 337, 336, 250, 249, 236. Found: 337.1907. $\text{C}_{21}\text{H}_{24}\text{N}_2\text{O}_2$ requires 337.1916.

3.2.20. 1-Methyl-7-phenylmelatonin (*N*-acetyl 1-methyl-5-methoxy-7-phenyltryptamine) (9a). Sodium hydride (11 mg, 0.27 mmol) was added to a solution of **8a** (38 mg, 0.12 mmol) in dry THF (1.0 mL) at 0 °C. The mixture was stirred for 30 min and iodomethane (15 μL , 0.25 mmol) was added and the mixture stirred for a further 2 h at room temperature. It was then quenched with water (2.0 mL), EtOAc (2 mL) and the aqueous layer extracted with EtOAc (3 \times 2 mL). The combined organic layers were washed with brine (2 mL), dried (MgSO_4) and concentrated under reduced pressure. The residue was purified by column chromatography (SiO_2 , EtOAc) followed by preparative HPLC (Chiralpak AD, 20 \times 250, 10 Å, 10 mL min^{-1} , hexane/IPA 80:20, 254 nm, 1 mL loop, retention time = 11.8 min) to give **9a**, 0.023 g (0.071 mmol, 59%), white crystals, mp

111 °C; ^1H NMR (CD_2Cl_2) δ 1.85 (s, 3H), 2.85 (t, $J = 6.8$ Hz, 2H), 3.15 (s, 3H), 3.47 (m, $J = 6.8$ Hz, 2H), 3.79 (s, 3H), 5.69 (br s, 1H), 6.61 (d, $J = 2.5$ Hz, 1H), 6.75 (s, 1H), 6.98 (d, $J = 2.5$ Hz, 1H) 7.37–7.34 (m, 5H); ^{13}C NMR (CD_2Cl_2) δ 24.9, 27.0, 38.1, 41.6, 57.5, 101.5, 112.8, 115.7, 129.1, 129.5, 129.6, 129.6, 131.3, 131.5, 131.7, 141.7, 154.9, 171.5; MS *m/e* 322 (23%, M^+), 263, 250 (100). Found: M^+ 322.1677. $\text{C}_{20}\text{H}_{22}\text{N}_2\text{O}_2$ requires 322.1681.

3.2.21. *N*-Propanoyl 5-methoxy-1-methyl-7-phenyltryptamine (9b). 0.0313 g, 0.0932 mmol, 73%, white crystals, mp 109 °C (CH_2Cl_2); ^1H NMR (CD_2Cl_2) δ 1.10 (t, $J = 5.7$ Hz, 3H), 2.14 (q, $J = 5.7$ Hz, 2H), 2.91 (t, $J = 5.1$ Hz, 2H), 3.21 (s, 3H), 3.54 (q, $J = 4.9$ Hz, 2H), 3.85 (s, 3H), 5.64 (br s, 1H), 6.67 (d, $J = 1.9$ Hz, 1H), 6.80 (s, 1H), 7.03 (d, $J = 1.9$ Hz, 1H), 7.38–7.43 (m, 5H); ^{13}C NMR (CD_2Cl_2) δ 11.5, 27.0, 31.5, 38.1, 41.4, 57.5, 101.5, 112.9, 115.7, 129.1, 129.5, 129.6, 131.3, 131.5, 131.7, 141.7, 154.9, 175.1; MS *m/e* 336, 263, 250 (100). Found: 336.1831. $\text{C}_{21}\text{H}_{24}\text{N}_2\text{O}_2$ requires 336.1838.

3.2.22. *N*-Butanoyl 5-methoxy-1-methyl-7-phenyltryptamine (9c). 0.0251 g, 0.072 mmol, 69%, white crystals, mp 109 °C (CH_2Cl_2); ^1H NMR δ 0.92 (t, $J = 5.5$ Hz, 3H), 1.61 (m, 2H), 2.09 (t, $J = 5.6$ Hz, 2H), 2.92 (t, $J = 5.1$ Hz, 2H), 3.21 (s, 3H), 3.55 (q, $J = 4.9$ Hz, 2H), 3.86 (s, 3H), 5.64 (br s, 1H), 6.67 (d, $J = 1.8$ Hz, 1H), 6.80 (s, 1H), 7.04 (d, $J = 1.8$ Hz, 1H), 7.39–7.44 m, 5H; ^{13}C NMR (CD_2Cl_2) δ 15.33, 20.9, 27.1, 38.1, 40.4, 41.4, 57.6, 101.6, 112.9, 115.7, 129.1, 129.5, 129.6, 131.3, 131.5, 131.7, 131.8, 141.7, 155.0, 174.3; MS *m/e* 350, 263, 250 (100). Found: 350.1995. $\text{C}_{22}\text{H}_{26}\text{N}_2\text{O}_2$ requires 350.1994.

3.3. Pharmacology

The affinity of the analogues was determined in competition radioligand binding assays using 2- ^{125}I iodomelatonin (specific activity 2000 Ci/mol, GE Healthcare, UK) as described previously²⁸ on the recombinant human MT_1 and MT_2 receptor subtypes expressed in NIH 3T3 cells, kindly supplied by Dr. S. M. Reppert (Harvard Medical School, Boston, MA). The biological activity of the analogues was assessed in a well established, specific model of melatonin action, the pigment aggregation response of *X. laevis* melanophores.¹ In these cells, melatonin triggers a translocation of pigment, normally distributed throughout the cell, towards the cell centre. This response is termed pigment aggregation and is quantified by measuring the change in light (630 nm) absorbance of the cells as the pigment concentrates near the cell centre. In the present study, a clonal melanophore line, generously provided by Dr. Michael Lerner (Department of Dermatology, University of Texas), was used.

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References and notes

- Sugden, D.; Davidson, K.; Hough, K. T.; Teh, M.-T. *Pigment Cell Res.* **2004**, *17*, 454–460.
- Zlotos, D. P. *Arch. Pharm. Chem. Life Sci.* **2005**, *338*, 229–247.
- Ebisawa, T.; Karne, S.; Lerner, M. R.; Reppert, S. M. *Proc. Natl. Acad. Sci. U.S.A.* **1994**, *91*, 6133–6137.
- Reppert, S. M.; Weaver, D. R.; Ebisawa, T. *Neuron* **1994**, *13*, 1177–1185.
- Reppert, S. M.; Weaver, D. R.; Cassone, V. M.; Godson, C.; Kolakowski, L. F. *Neuron* **1995**, *15*, 1003–1015.
- Reppert, S. M.; Weaver, D. R.; Godson, C. *TIPS* **1996**, *17*, 100–102.
- Barrett, P.; Conway, S.; Morgan, P. J. *J. Pineal Res.* **2003**, *35*, 221–230.
- Dubocovich, M. L.; Masana, M. I.; Jacob, S.; Sauri, D. M. *Naunyn Schmiedebergs Arch. Pharmacol.* **1997**, *355*, 365–375.
- Sugden, D.; Pickering, H.; Teh, M.-T.; Garratt, P. J. *Biol. Cell* **1997**, *89*, 531–537.
- Faust, R.; Garratt, P. J.; Jones, R.; Yeh, L.-K.; Tsotinis, A.; Panoussopoulou, M.; Calogeropoulou, T. P.; Teh, M.-T.; Sugden, D. *J. Med. Chem.* **2000**, *43*, 1050–1061.
- Spadoni, G.; Balsamini, C.; Bedini, A.; Carey, A.; Diamantini, G.; DiGiacomo, B.; Tontini, A.; Tarzia, G.; Nonno, R.; Lucini, V.; Pannacci, M.; Stankov, B. M.; Fraschini, F. *Med. Chem. Res.* **1998**, *8*, 487–498.
- Spadoni, G.; Balsamini, C.; Diamantini, G.; Tontini, A.; Tarzia, G.; Mor, M.; Rivara, S.; Plazzi, P. V.; Nonno, R.; Lucini, V.; Pannacci, M.; Fraschini, F.; Stankov, B. M. *J. Med. Chem.* **2001**, *44*, 2900–2912.
- Descamps-François, C.; Yous, S.; Chavatte, P.; Audinot, V.; Bonnaud, A.; Boutin, J. A.; Delagrangé, P.; Bennejean, C.; Renard, P.; Lesieur, D. *J. Med. Chem.* **2003**, *46*, 1127–1129.
- Audinot, V.; Mailliet, F.; Lahaye-Brasseur, C.; Bonnaud, A.; Le Gall, A.; Amossé, C.; Dromaint, S.; Rodriguez, M.; Nagel, N.; Galizzi, J.-P.; Malpoux, B.; Guillaumet, G.; Lesieur, D.; Lefoulon, F.; Renard, P.; Delagrangé, P.; Boutin, J. A. *Naunyn Schmiedebergs Arch. Pharmacol.* **2003**, *367*, 553–561.
- Epperson, J. R.; Deskus, J. A.; Gentile, A. J.; Iben, L.; Ryan, E.; Sarbin, N. S. *Biol. Med. Chem. Lett.* **2004**, *14*, 1023–1026.
- Rivara, S.; Mor, M.; Silva, C.; Zuliani, V.; Vacondio, F.; Spadoni, G.; Bedini, A.; Tarzia, G.; Lucini, V.; Pannacci, M.; Fraschini, F.; Plazzi, P. V. *J. Med. Chem.* **2003**, *46*, 1429–1439.
- Levoye, A.; Jockers, R.; Ayoub, M. A.; Delagrangé, P.; Savaskan, E.; Guillaume, J. L. *Chronobiol. Int.* **2006**, *23*, 419–426.
- Mor, M.; Rivara, S.; Silva, C.; Bordini, F.; Plazzi, P. V.; Spadoni, G.; Diamantini, G.; Balsamini, C.; Tarzia, G.; Fraschini, F.; Lucini, V.; Nonno, R.; Stankov, B. M. *J. Med. Chem.* **1998**, *41*, 3831–3844.
- Spadoni, G.; Mor, M.; Tarzia, G. *Biol. Signals Receptors* **1999**, *8*, 15–23.
- Leclerc, V.; Yous, S.; Delagrangé, P.; Boutin, J. A.; Renard, P.; Lesieur, D. *J. Med. Chem.* **2002**, *45*, 1853–1859.
- Tsotinis, A.; Vlachou, M.; Papahadjis, D. P.; Calogeropoulou, T.; Nikas, S.; Garratt, P. J.; Piccio, V.; Vonhoff, S.; Davidson, K.; Teh, M.-T.; Sugden, D. *J. Med. Chem.* **2006**, *49*, 3509–3519.
- Somei, M.; Hatterli, A.; Suzuki, N. *PCT Int. Appl.* **2005**.
- Büchi, G.; Mak, C.-P. *J. Org. Chem.* **1977**, *42*, 1784–1786.
- Engler, T. A.; Furness, K.; Malhotra, S.; Sanchez-Martinez, C.; Shih, C.; Xie, W.; Zhu, G.; Zhou, X.; Conner, S.

- Faul, M. M.; Sullivan, K. A.; Kollis, S. P.; Brooks, H. B.; Patel, B.; Schultz, R. M.; DeHahn, T. B.; Kirmani, K.; Spencer, C. D.; Watkins, S. A.; Considine, E. L.; Dempsey, J. A.; Ogg, C. A.; Stamm, N. B.; Anderson, B. A.; Campbell, R. M.; Vasudevan, V.; Lytle, M. L. *Biol. Med. Chem. Lett.* **2003**, *13*, 2261–2267.
25. Suzuki, A. *J. Organomet. Chem.* **1999**, *576*, 147–168.
26. Glennon, R. A.; Schubert, E.; Jacyno, J. M.; Rosecrans, J. A. *J. Med. Chem.* **1980**, *23*, 1222–1226.
27. Bartoli, G.; Palmieri, G.; Bosco, M.; Dalpozzo, R. *Tetrahedron Lett.* **1989**, *30*, 2129–2132.
28. Teh, M. T.; Sugden, D. *Br. J. Pharm.* **1998**, *125*, 75P.